JB commentary

An impact of CCN2–BMP-2 complex upon chondrocyte biology: evoking a signalling pathway bypasses ERK and Smads?

Received June 2, 2011; accepted June 27, 2011; published online July 18, 2011

Shingo Maeda*

Department of Medical Joint Materials, Graduate School of Medical and Dental Sciences, Kagoshima University, 8-35-1 Sakuragaoka, Kagoshima 890-8544, Japan

*Shingo Maeda, MD, PhD, Department of Medical Joint Materials, Graduate School of Medical and Dental Sciences, Kagoshima University, 8-35-1 Sakuragaoka, Kagoshima 890-8544, Japan. Tel: +81-99-275-5381, Fax: +81-99-265-4699, email: s-maeda@m3.kufm.kagoshima-u.ac.jp

CCN family 2/connective tissue growth factor (CCN2/ CTGF) is a secreted protein that regulates diverse cellular functions. In addition to being a growth factor to transmit mitogen-activated protein kinase (MAPK) signalling into cells, through largely unknown mechanism, CCN2 antagonizes bone morphogenetic proteins (BMPs) by direct interaction. CCN2 and BMPs co-localize in cartilage, and both of them promote proliferation and differentiation of chondrocytes in vivo and in vitro. However, it was unclear whether these growth factors act cooperatively, or mutually inhibitory in chondrocyte biology. In addition, an information whether the heterooligomer of CCN2 and BMPs has any physiological roles in skeletogenesis was completely missing. Takigawa and his colleagues have recently reported that CCN2 and BMP-2 interacted directly to mutually interfere with their downstream ERK and Smad signalling, whereas the CCN2-BMP-2 complex promoted chondrocyte differentiation [Maeda et al. (2009) J. Biochem. 145, 207–216]. This contradictory finding shed light on a possibility that the complex of CCN2 and BMP-2 is capable of activating an additional non-canonical signalling pathway to promote chondrocyte differentiation.

Keywords: BMP-2/CCN2/chondrocytes/ERK/Smad.

Abbreviations: BMP, bone morphogenetic protein; TGF- β , transforming growth factor- β ; MAPK, mitogen-activated protein kinase; ERK, extracellular signal-regulated kinase.

Members of the CCN family of cysteine-rich (CR) secreted protein include cysteine-rich protein 61 (Cyr61)/ CCN1, connective tissue growth factor (CTGF)/CCN2, nephroblastoma over-expressed gene (Nov)/CCN3,



CCN proteins are highly conserved and contain four distinct structural modules: an amino-terminal insulinlike growth factor-binding domain (IGFB), followed by a CR/von Willebrand type C domain-containing domain, a thrombospondin type 1 repeat (TSP1) and a carboxyl-terminal cystine knot (CT) domain (Fig. 1A). All of these modules are highly interactive with many other molecules such as aggrecan (6), fibronectins (7), transforming growth factor-\u00b31 (TGF-\u00b31) (8), BMP-4 (8) and BMP-7 (9). CR module is similar to those found in a BMP antagonist chordin, and is responsible for binding to BMPs (10). BMPs are member of TGF-ß family, which bind to two distinct Type II and Type I serine/threonine kinase receptors, which in turn phosphorylate receptor-regulated Smads (Smad1, 5 and 8 for BMP signalling) to transduce canonical BMP signalling (Fig. 1B) (11). BMPs promote proliferation and differentiation of chondrocytes through Smad1/5/8 (12). Binding of CCN2 to BMP-4 inhibited interaction of BMP-4 with BMP receptors, thereby inhibited BMP signalling (8). Other CCN family proteins also bind to BMPs to inhibit BMP signalling. For instance, Nov/CCN3 binds to BMP-2 (13), WISP1/CCN4 binds to BMP-2 (14) and WISP3/ CCN6 interacts with BMP-4 (15). Among them, interestingly, only WISP-1/CCN4 was not inhibitory to BMP signalling, rather promoted BMP-2 activity by inducing α_5 -integrin expression to enhance the accessibility of BMP-2 for bone marrow stroma cells, thereby accelerate bone formation to increase bone volume of the transgenic mice skeleton (14). These findings indicate that CCN family proteins can serve as BMP antagonists or agonists in context-dependent manner.



Fig. 1 Signal cross-talk between CCN2 and BMP-2 in chondrocyte proliferation and differentiation. (A) Structural domains of CCN2 protein. IGFB, CR/ VWC, TSP1 repeat and CT domains are shown. (B) Combined application of CCN2 and BMP-2 inhibits BMP-Smad pathway, BMP-ERK pathway, CCN2-ERK pathway and chondrocyte proliferation, whereas promotes chondrocyte differentiation.

In case of Noggin, a representative BMP antagonist, the knock-out mice showed accelerated BMP signalling, which resulted in overgrowth of skeletal elements (16). On the other hand, CCN2-deficient mice showed an opposite skeletal phenotype of chondrodysplasia (5). These evidences from mouse genetics suggest that a contribution of CCN2 as a BMP antagonist in chondrogenesis is dispensable, or it is possible that CCN2–BMPs complex have an unidentified positive function in cartilage development.

Recently, Takigawa and his colleague (17) studied a possible interaction between CCN2 and BMP-2, and the functional impacts of the resulting heterooligomer on chondrocytes. CCN2 interacted with BMP-2 with a dissociation constant (K_D) of 0.77 nM, while CCN2 had been reported to bind to BMP-4 at a $K_{\rm D}$ of 5 nM (8), and to BMP-7 with a K_D of 14 nM (9). Not only CR module of CCN2, but also modules of IGFB and CT interacted to BMP-2, among which CT module showed the highest affinity (17). Together with the fact that cysteine residues in CR module of CCN2 are 80% homologous to those found in chordin (10), BMP-2 is likely to have the strongest affinity to CCN2 among these osteogenic BMPs. As expected, combined application of CCN2 and BMP-2 inhibited both BMP-2-induced phosphorylation of Smad1/5/8 and

proliferation of HCS-2/8 (17). This was probably achieved though preventing BMP-2 from interacting with BMP receptors, as a case for BMP-4 (Fig. 1B) (8). In addition to Smad pathway, BMPs transmit signals through non-Smad pathways, including extracellular signal-regulated kinase (ERK), JNK and p38 mitogen-activated protein kinase (MAPK) pathways (Fig. 1B) (18). Co-treatment of CCN2 abolished BMP-2-induced phosphorylation of ERK1/2, whereas did not affect that of p38 (17). CCN2 activates ERK pathway and JNK pathway to up-regulate proliferation of chondrocytes, and p38 pathway to promote differentiation of HCS-2/8 cells (Fig. 1B) (19, 20). Interestingly, addition of BMP-2 dramatically suppressed CCN2-induced phosphorylation of ERK1/2, but not that of p38, which in turn completely cancelled the accelerated proliferation of HCS-2/8 cells by CCN2 (17). Surprisingly, however, combined treatment of BMP-2 and CCN2 not only showed no inhibition against the promoted chondrocyte differentiation by either factor alone, but also further enhanced the production of cartilage extracellular matrix proteins (17). Because the phosphorylation level of p38, responsible for chondrocyte differentiation, was unchanged by BMP-2–CCN2 co-application as mentioned above, this collaborative effect by these two growth factors on chondrocyte differentiation seemed independent of neither Smad pathway nor p38 MAPK cascade. It should be noted that the effect of combined treatment of BMP-2 and CCN2 on chondrocyte differentiation was 'additive' rather than 'synergistic', therefore, it is not clear whether the complex formation by BMP-2 and CCN2 was indispensable in this context, or not. To date, possible physiological receptors for CCN2 responsible for promoting cell proliferation and differentiation are largely unknown. It is possible that the heterooligomers of BMPs and CCN family proteins bind to some receptors belonging to other signalling systems to evoke an alternative non-canonical signal cascade into certain cells (Fig. 1B).

Funding

Grant-in-Aid for Scientific Research (KAKENHI)(C)(23592221).

Conflict of interest

None declared.

References

- 1. Leask, A. and Abraham, D.J. (2006) All in the CCN family: essential matricellular signaling modulators emerge from the bunker. *J. Cell Sci.* **119**, 4803–4810
- Kronenberg, H.M. (2003) Developmental regulation of the growth plate. *Nature* 423, 332–326
- Nakanishi, T., Kimura, Y., Tamura, T., Ichikawa, H., Yamaai, Y., Sugimoto, T., and Takigawa, M. (1997) Cloning of a mRNA preferentially expressed in chondrocytes by differential display-PCR from a human chondrocytic cell line that is identical with connective tissue growth factor (CTGF) mRNA. *Biochem. Biophys. Res. Commun.* 234, 206–210
- Nakanishi, T., Nishida, T., Shimo, T., Kobayashi, K., Kubo, T., Tamatani, T., Tezuka, K., and Takigawa, M. (2000) Effects of CTGF/Hcs24, a product of a hypertrophic chondrocyte-specific gene, on the proliferation and differentiation of chondrocytes in culture. *Endocrinology* 141, 264–273
- Ivkovic, S., Yoon, B.S., Popoff, S.N., Safadi, F.F., Libuda, D.E., Stephenson, R.C., Daluiski, A., and Lyons, K.M. (2003) Connective tissue growth factor coordinates chondrogenesis and angiogenesis during skeletal development. *Development* 130, 2779–2791
- Aoyama, E., Hattori, T., Hoshijima, M., Araki, D., Nishida, T., Kubota, S., and Takigawa, M. (2009) N-terminal domains of CCN family 2/connective tissue growth factor bind to aggrecan. *Biochem. J.* 420, 413–420
- Chen, Y., Abraham, D.J., Shi-Wen, X., Pearson, J.D., Black, C.M., Lyons, K.M., and Leask, A. (2004) CCN2 (connective tissue growth factor) promotes fibroblast adhesion to fibronectin. *Mol. Biol. Cell.* 15, 5635–5646
- 8. Abreu, J.G., Ketpura, N.I., Reversade, B., and De Robertis, E.M. (2002) Connective-tissue growth factor

Signal cross-talk between CCN2 and BMP-2 in chondrocytes

(CTGF) modulates cell signalling by BMP and TGF-β. *Nat. Cell Biol.* **4**, 599–604

- Nguyen, T.Q., Roestenberg, P., van Nieuwenhoven, F.A., Bovenschen, N., Li, Z., Xu, L., Oliver, N., Aten, J., Joles, J.A., Vial, C., Brandan, E., Lyons, K.M., and Goldschmeding, R. (2008) CTGF inhibits BMP-7 signaling in diabetic nephropathy. J. Am. Soc. Nephrol. 19, 2098–2107
- Inkson, C.A., Ono, M., Kuznetsov, S.A., Fisher, L.W., Robey, P.G., and Young, M.F. (2008) TGF-β1 and WISP-1/CCN-4 can regulate each other's activity to cooperatively control osteoblast function. *J. Cell. Biochem.* 104, 1865–1878
- Miyazono, K., Kamiya, Y., and Morikawa, M. (2010) Bone morphogenetic protein receptors and signal transduction. J. Biochem. 147, 35–51
- Retting, K.N., Song, B., Yoon, B.S., and Lyons, K.M. (2009) BMP canonical Smad signaling through Smad1 and Smad5 is required for endochondral bone formation. *Development* 136, 1093–1104
- Minamizato, T., Sakamoto, K., Liu, T., Kokubo, H., Katsube, K., Perbal, B., Nakamura, S., and Yamaguchi, A. (2007) CCN3/NOV inhibits BMP-2induced osteoblast differentiation by interacting with BMP and Notch signaling pathways. *Biochem. Biophys. Res. Commun.* 354, 567–573
- Ono, M., Inkson, C.A., Kilts, T.M., and Young, M.F. (2011) WISP-1/CCN4 regulates osteogenesis by enhancing BMP-2 activity. J. Bone Miner. Res. 26, 193–208
- Nakamura, Y., Weidinger, G., Liang, J.O., Aquilina-Beck, A., Tamai, K., Moon, R.T., and Warman, M.L. (2007) The CCN family member Wisp3, mutant in progressive pseudorheumatoid dysplasia, modulates BMP and Wnt signaling. J. Clin. Invest. 117, 3075–3086
- Brunet, L.J., McMahon, J.A., McMahon, A.P., and Harland, R.M. (1998) Noggin, cartilage morphogenesis, and joint formation in the mammalian skeleton. *Science* 280, 1455–1457
- Maeda, A., Nishida, T., Aoyama, E., Kubota, S., Lyons, K.M., Kuboki, T., and Takigawa, M. (2009) CCN family 2/connective tissue growth factor modulates BMP signalling as a signal conductor, which action regulates the proliferation and differentiation of chondrocytes. *J. Biochem.* 145, 207–216
- Derynck, R. and Zhang, Y.E. (2003) Smad-dependent and Smad-independent pathways in TGF-β family signalling. *Nature* 425, 577–584
- Yosimichi, G., Nakanishi, T., Nishida, T., Hattori, T., Takano-Yamamoto, T., and Takigawa, M. (2001) CTGF/Hcs24 induces chondrocyte differentiation through a p38 mitogen-activated protein kinase (p38MAPK), and proliferation through a p44/42 MAPK/extracellular-signal regulated kinase (ERK). *Eur. J. Biochem.* 268, 6058–6065
- 20. Kubota, S., Kawaki, H., Kondo, S., Yosimichi, G., Minato, M., Nishida, T., Hanagata, H., Miyauchi, A., and Takigawa, M. (2006) Multiple activation of mitogen-activated protein kinases by purified independent CCN2 modules in vascular endothelial cells and chondrocytes in culture. *Biochimie* 88, 1973–1981